



# A 2x2 factorial microarray experiment

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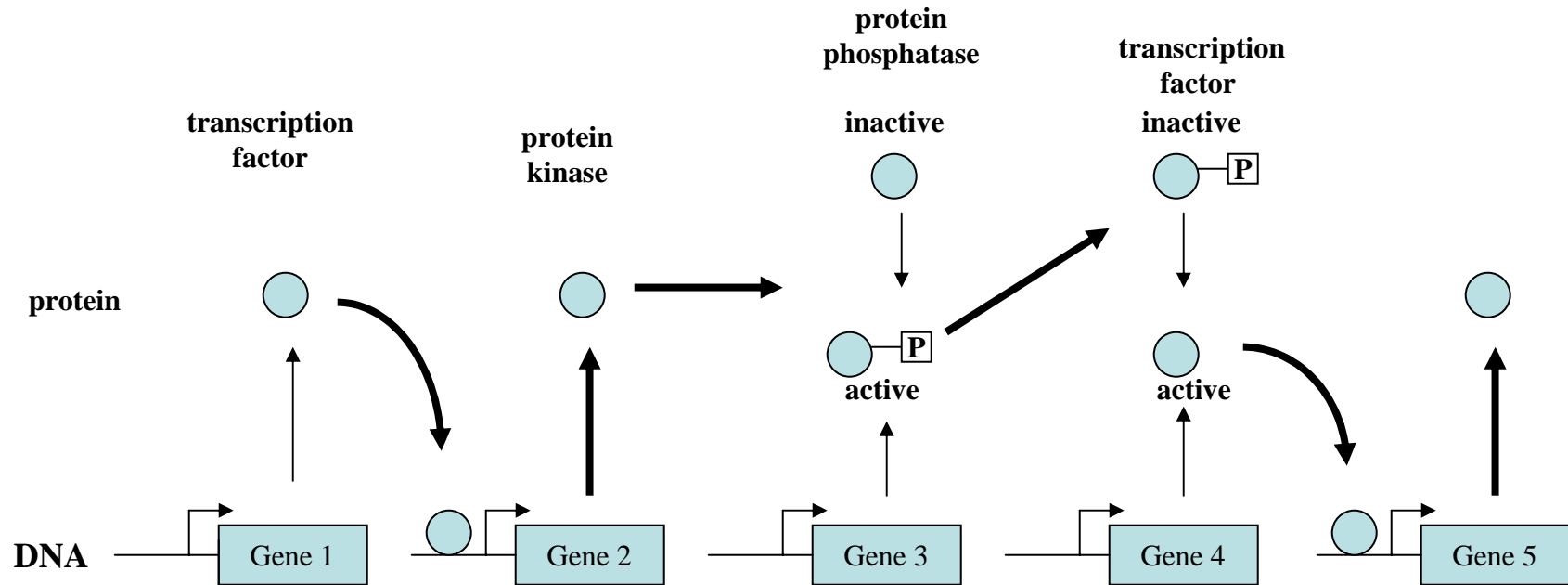
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# Complexity of genomic data

- The functioning of cells is a complex and highly structured process.
- In the next slide we show a stylized [biochemical pathway](#) (adapted from Wagner, 2001).
- There are transcription factors, protein kinase and protein phosphatase reactions.
- Tools are being developed that allow us to explore this functioning in a multitude of different ways.

# An example of the interactions between some genes (adapted from Wagner 2001)



# Overview

- Wagner (2001) suggests that the holy grail of functional genomics is the reconstruction of **genetic networks**.
- In this tutorial we examine some methods for doing this in **factorial genome wide RNA expression experiments**.
- Such experiments are easy to carry out and are becoming widespread. Tools for analyzing them are badly needed.

# Gene effects

- A factor can either inhibit or enhance the production of mRNA for any gene.
- The inhibition or enhancement of mRNA production for any given gene can affect transcription for other genes either through inhibition or enhancement.

# Targets

- We define a **target** of a factor to be a gene whose expression of mRNA is altered by the presence of the factor.
- A **primary target** is a target that is directly affected by the factor.
- A **secondary target** is a target whose transcription is altered only via the effects of some other genes, i.e., can be traced back to one or more primary targets.

# CX experiment

- There are two factors
  - Estrogen, E: known to affect transcription of various genes (some known, some unknown).
  - Cyclohexamide, CX: known to stop all translation (with very few exceptions).
- The design is a classical **2x2 factorial design**, with two replicates.
- We are interested in the main effects and interactions for E and CX.

# CX experiment

- We identify as **targets** all genes whose expression of mRNA is affected by the application of E.
- A target can be either primary or secondary
  - **primary** if E directly affects expression of mRNA.
  - **secondary** if mRNA production is affected by some other gene and can be traced back to a primary target.



# Scenario 1

- Assume that there are two related genes, B and D, where
  - B is a primary target of E,
  - D is a secondary target only via B.
- Neither is expressed initially.
- E causes B to be expressed and this in turn causes D to be expressed.
- The addition of CX by itself may not affect expression of either B or D.

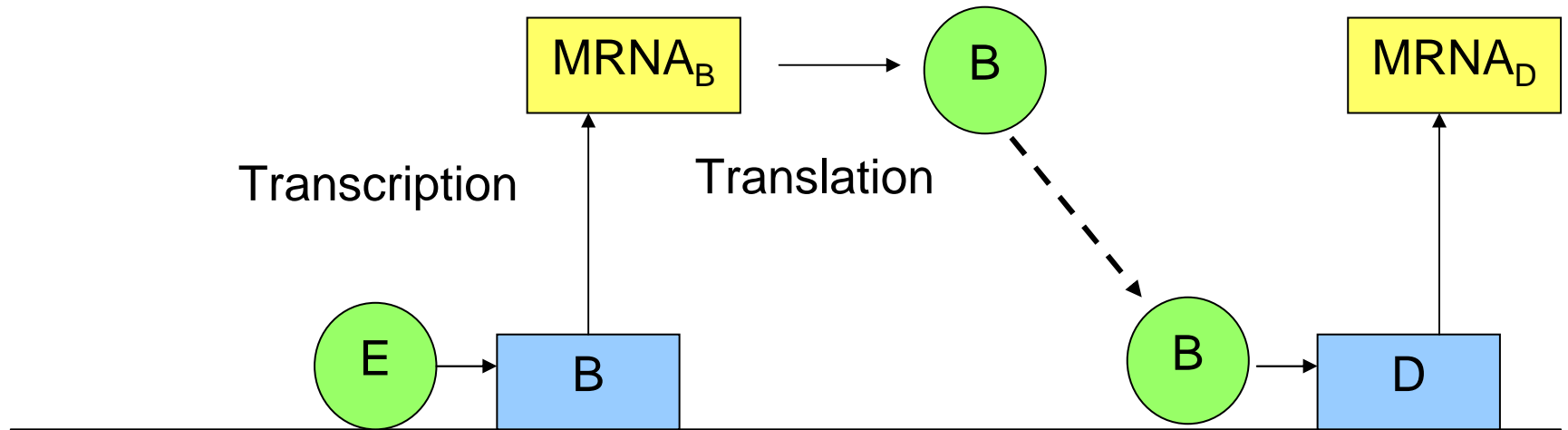
No factors applied



Gene B is not active

Gene D is not active

E only



B is a Primary Target of E

D is a Secondary Target of E

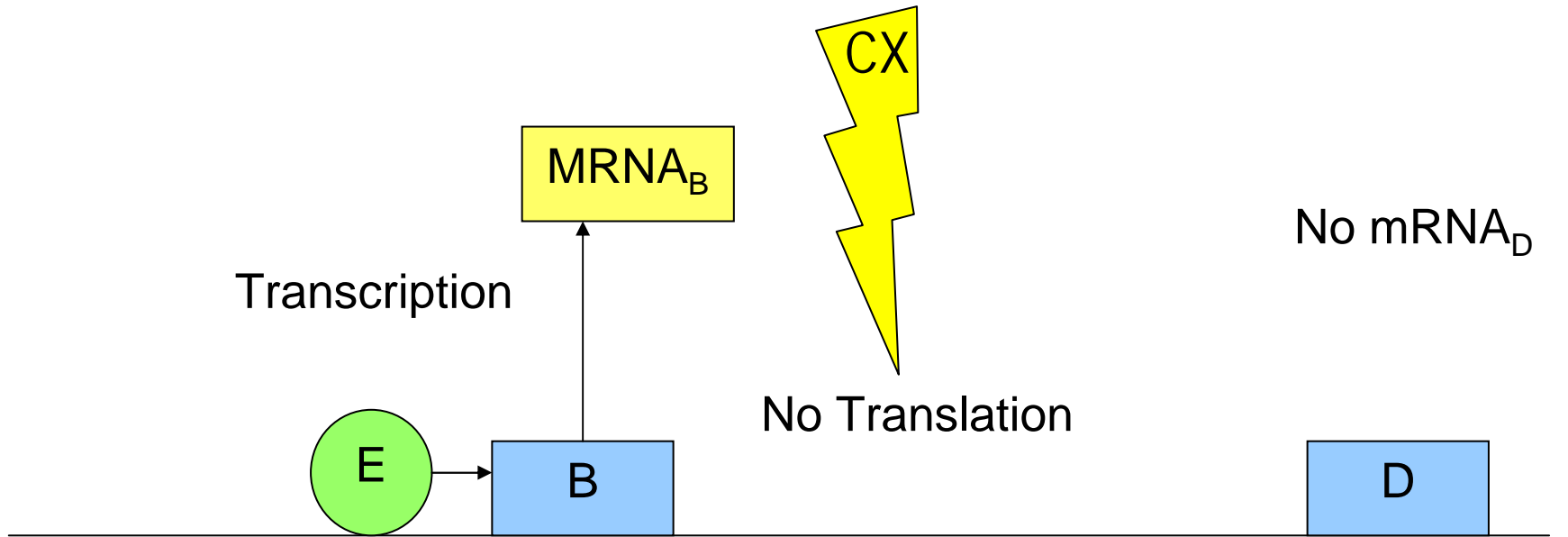
Production of mRNA<sub>B</sub> is enhanced by E

Production of mRNA<sub>D</sub> is enhanced by B

# Scenario 1

- In the presence of both CX and E we see increased expression of mRNA<sub>B</sub> but not of mRNA<sub>D</sub>.
- CX stops translation of B and hence transcription of D.
- This will be one of the principles we can use to differentiate between primary targets of E (such as B) and secondary targets of E (such as D).

E and CX both present



B is a Primary Target

Production of mRNA<sub>B</sub> is enhanced by E

Production of mRNA<sub>D</sub> is decreased (prevented)

# Interpretation: Scenario 1

	mRNA <sub>B</sub>	mRNA <sub>D</sub>
Nothing	Low	Low
E	High	High
CX	Low(?)	Low (?)
E and CX	High	Low

# Scenario 1

- Note that while we show a direct relationship between the expression of B and of D we cannot detect such a relationship from these data (the purpose of this scenario is purely pedagogical).
- Other scenarios include
  - Suppression of D by B, enhancement of B by E.
  - Enhancement of D by B, and suppression of B by E.

# CX experiment

- Assume the following linear model for the observed expression response (possibly on transformed data) of any given gene

$$y_{ig} = \mu_g + \beta_{Eg} x_{1i} + \beta_{CXg} x_{2i} + \beta_{E:CX,g} x_{1i} x_{2i} + \varepsilon_{ig}$$

- $i$  indexes chips and  $g$  indexes genes.
- $x_1$  indicates the presence of E and  $x_2$  indicates the presence of CX.



# Inference

- The 2x2 CX microarray experiment measures the expression response of each gene under each of the four factor combinations.
- But there is a difference, B is a primary target of E, while D is a secondary target of E.

# Inference

- If gene  $X$  is any target for  $E$ , the level of  $\text{mRNA}_X$  might not change when  $E$  is added.
- $\text{mRNA}_X$  might already be being made as fast as possible, so addition of  $E$  has no effect.
- Production of  $\text{mRNA}_X$  might already be suppressed by some other compound.
- A true baseline would help in resolving these situations.

# Inference

- The introduction of CX provides a form of baseline.
- Since (among other things) CX halts translation we should be able to use the presence or absence of CX to find out about primary versus secondary targets.

# Inference

- For any gene we can interpret the coefficients in the linear model as follows.
- The parameter  $\beta_E$  can be interpreted as the main effect of E.
- Genes for which  $\beta_E$  is different from zero are potential **targets**.
- As noted previously, not all targets will have  $\beta_E$  different from zero.

# Inference

- The parameter  $\beta_{CX}$  can be interpreted as the main effect of CX.
- If  $\beta_{CX}$  is different from zero, this suggests that production of mRNA is **translationally regulated**.
- The interpretation of the interaction  $\beta_{E: CX}$  is more difficult.

# Primary targets

- Consider the case where we have only CX and CX+E.
- Since CX halts all translation, then any differences between the condition where CX alone is present and CX+E is present should indicate primary targets of E.
- This is equivalent to testing the hypothesis
$$H_0: \mu + \beta_E + \beta_{CX} + \beta_{E: CX} = \mu + \beta_{CX}, \text{ i.e.,}$$
$$H_0: \beta_E + \beta_{E: CX} = 0$$

# Primary targets

- Genes for which the hypothesis

$$H_0: \mu + \beta_E + \beta_{CX} + \beta_{E:CX} = \mu + \beta_{CX}$$

is rejected are candidates for **primary targets**.

- Those with  $\beta_E$  different from zero, but for which we do not reject  $H_0$ , are **secondary targets**.
- It seems likely that some inference may be drawn from the relationship between  $\beta_E$  and  $\beta_{E:CX}$ , their signs and their significance levels.

# Scenario 1

	Primary	Secondary
$\beta_E$	$> 0$	$> 0$
$\beta_{CX}$	$= 0$	$= 0$
$\beta_{E:CX}$	$= 0$	$-\beta_E$



# Limitations

- While we may identify genes that are potentially primary targets and those that are potentially secondary targets we cannot identify gene—gene interactions, or feedback loops.
- We can observe the effects but not attribute them.
- The use of relevant metadata, biological and publication, seems pertinent and could help resolve some of the interactions.