

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).



## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```



```

[1] 5

> which(myresult$permutation_p<=0.05)

[1] 93 101 247 790 946

> sum(myresult$bootstrap_p<=0.05)

[1] 10

> which(myresult$bootstrap_p<=0.05)

[1] 239 638 662 669 741 790 795 927 931 946

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 40

> which(myresult2$bootstrap_p<=0.05)

[1] 24 41 44 59 131 143 173 223 253 293 296 311 333 336 340 347 368 387 458
[20] 481 486 491 540 544 573 597 606 611 617 641 668 672 713 733 738 798 888 907
[39] 938 960

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 5

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.



```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 57

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 52

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 52

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 24 32 33 51 53 63 70 93 106 110 113 126 127 135 140 166 169 200 214
[20] 231 241 263 288 315 386 399 400 437 454 479 480 495 572 632 665 681 689 691
[39] 705 735 747 750 760 781 798 814 823 826 842 867 871 883 901 903 930 949 951

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 24 32 33 51 53 63 93 110 113 127 135 140 159 166 169 200 231 240 241
[20] 263 288 289 295 315 386 399 400 437 454 480 495 607 648 665 681 689 691 735
[39] 747 750 781 798 814 823 827 842 883 899 903 930 949 951

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 3 24 32 33 51 55 63 93 110 113 126 127 140 159 166 169 241 263 288
[20] 295 315 386 399 400 437 454 480 495 608 665 681 689 705 735 747 760 781 798
[39] 814 819 823 826 827 842 871 883 899 901 903 930 949 951

> con1_adj_p <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adj_p<=0.05/3)

[1] 10

```



```

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 5

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 7

> which(con2_adjp<=0.05/3)

[1] 93 127 263 495 689

> which(con3_adjp<=0.05/3)

[1] 32 93 166 288 480 495 681

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1  3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p   3000   -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 62

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 66

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 61

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 4 63 92 113 117 131 140 142 177 182 197 218 236 237 245 253 284 291 292
[20] 324 335 339 350 363 368 371 390 396 414 416 428 448 469 483 500 512 522 540
[39] 562 582 584 594 599 606 609 647 655 669 672 697 705 817 821 839 846 853 872
[58] 881 961 974 989 996

```



```

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 4 18 48 80 92 113 124 131 140 142 177 182 197 230 236 237 253 284 291
[20] 292 327 339 350 363 368 390 414 416 428 434 435 448 469 483 500 508 512 522
[39] 540 562 582 584 594 599 605 609 619 647 652 655 669 672 697 714 734 744 817
[58] 839 846 853 881 950 961 974 989 996

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 4 18 48 113 124 131 140 142 177 182 197 218 236 237 245 253 284 291 292
[20] 324 335 339 360 371 385 396 416 428 435 448 469 483 500 508 512 518 522 540
[39] 553 562 582 584 594 599 605 609 647 655 672 697 700 714 744 846 853 881 950
[58] 961 974 989 996

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 6

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 8

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 5

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the gemone-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

[1] "/private/var/folders/r0/l4fjk6cj5xj0j3brt4bplpl40000gt/T/RtmpsUgICl/Rinst11be84d441b5a/RBM/"

```



```

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

      IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1   Min.    :0.01058   Min.    :0.01187   Min.    :0.009103
cg00002426: 1   1st Qu.:0.04111   1st Qu.:0.04407   1st Qu.:0.041543
cg00003994: 1   Median :0.08284   Median :0.09531   Median :0.087042
cg00005847: 1   Mean    :0.27397   Mean    :0.28872   Mean    :0.283729
cg00006414: 1   3rd Qu.:0.52135   3rd Qu.:0.59031   3rd Qu.:0.558575
cg00007981: 1   Max.    :0.97069   Max.    :0.96937   Max.    :0.970155
(Other)      :994                      NA's     :4
exmdata4[, 2]      exmdata5[, 2]      exmdata6[, 2]      exmdata7[, 2]
Min.    :0.01019   Min.    :0.01108   Min.    :0.01937   Min.    :0.01278
1st Qu.:0.04092   1st Qu.:0.04059   1st Qu.:0.05060   1st Qu.:0.04260
Median :0.09042   Median :0.08527   Median :0.09502   Median :0.09362
Mean    :0.28508   Mean    :0.28482   Mean    :0.27348   Mean    :0.27563
3rd Qu.:0.57502   3rd Qu.:0.57300   3rd Qu.:0.52099   3rd Qu.:0.52240
Max.    :0.96658   Max.    :0.97516   Max.    :0.96681   Max.    :0.95974
                      NA's     :1
exmdata8[, 2]
Min.    :0.01357
1st Qu.:0.04387
Median :0.09282
Mean    :0.28679
3rd Qu.:0.57217
Max.    :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t      1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0   1000  -none- numeric
ordfit_beta1   1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p    1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)

[1] 47

> sum(diff_results$permutation_p<=0.05)

[1] 54

```



```

> sum(diff_results$bootstrap_p<=0.05)

[1] NA

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 9

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] NA

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_perm)

```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
346	cg00331237	0.05972383	NA	0.08204769	0.08345662
677	cg00651216	0.06825629	0.12529090	0.14409190	0.13907250
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
927	cg00899659	0.10285880	0.22472200	0.22740700	0.23304960
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
19	0.80831380	0.73306440	0.82968340	0.84917800	
103	0.67349260	0.73510200	0.75715920	0.78981220	
346	0.05372019	0.06241126	0.06955040	0.09140985	
677	0.07669587	0.09597587	0.11690440	0.15194540	
764	0.90575890	0.88760470	0.90756300	0.90946790	
848	0.05230160	0.06119713	0.06542751	0.06240686	
851	0.50820240	0.34657470	0.66276570	0.64634510	
887	0.50259740	0.40111730	0.56646700	0.54552980	
927	0.12656770	0.10920420	0.15898110	0.25753470	
	diff_results\$ordfit_t[diff_list_perm]				
19	-2.547097				
103	-2.343784				



```

346          -3.328798
677          -3.457874
764          -1.560713
848          -1.687144
851          -2.986319
887          -3.368752
927          -2.533336
diff_results$permutation_p[diff_list_perm]
19          0
103         0
346         0
677         0
764         0
848         0
851         0
887         0
927         0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)

```

```

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
146 cg00134539 0.61101320    0.53321780    0.45999340    0.46787420
259 cg00234961 0.04192170    0.04321576    0.05707140    0.05327565
280 cg00260778 0.64319890    0.60488960    0.56735060    0.53150910
743 cg00717862 0.07999436    0.07873347    0.06089359    0.06171374
979 cg00945507 0.13432250    0.23854600    0.34749760    0.28903340
      exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
146    0.67191510    0.63137380    0.47929610    0.45428300
259    0.04030003    0.03996053    0.05086962    0.05445672
280    0.61920530    0.61925200    0.46753250    0.55632410
743    0.07594936    0.09062161    0.06475791    0.07271878
979    0.11848510    0.16653850    0.30718420    0.26624740
diff_results$ordfit_t[diff_list_boot]
146          5.636263
259         -2.833203
280          4.337628
743          2.918806
979         -4.968792
diff_results$bootstrap_p[diff_list_boot]
146          0
259          0
280          0
743          0
979          0

```