

# Package ‘CiteFuse’

May 1, 2024

**Type** Package

**Title** CiteFuse: multi-modal analysis of CITE-seq data

**Version** 1.17.0

**Description** CiteFuse package implements a suite of methods and tools for CITE-seq data from pre-processing to integrative analytics, including doublet detection, network-based modality integration, cell type clustering, differential RNA and protein expression analysis, ADT evaluation, ligand-receptor interaction analysis, and interactive web-based visualisation of the analyses.

**License** GPL-3

**Encoding** UTF-8

**Depends** R (>= 4.0)

**Imports** SingleCellExperiment (>= 1.8.0), SummarizedExperiment (>= 1.16.0), Matrix, mixtools, cowplot, ggplot2, gridExtra, grid, dbscan, uwot, Rtsne, S4Vectors (>= 0.24.0), igraph, scales, scran (>= 1.14.6), graphics, methods, stats, utils, reshape2, ggridges, randomForest, pheatmap, ggraph, grDevices, rhdf5, rlang, Rcpp, compositions

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**Author** Yingxin Lin [aut, cre],  
Hani Kim [aut]

**Maintainer** Yingxin Lin <yingxin.lin@sydney.edu.au>

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---

CiteFuse

*CiteFuse*

---

## Description

A function to runSNF for CITE seq data

**Usage**

```

CiteFuse(
  sce,
  altExp_name = "ADT",
  W_list = NULL,
  gene_select = TRUE,
  dist_cal_RNA = "correlation",
  dist_cal_ADT = "propr",
  ADT_subset = NULL,
  K_knn = 20,
  K_knn_Aff = 30,
  sigma = 0.45,
  t = 10,
  metadata_names = NULL,
  verbose = TRUE,
  topN = 2000
)

```

**Arguments**

sce	a SingleCellExperiment
altExp_name	expression name of ADT matrix
W_list	affinity list, if it is NULL, the function will calculate it.
gene_select	whether highly variable genes will be selected for RNA-seq to calculate similarity matrix using ‘scran’ package
dist_cal_RNA	similarity metrics used for RNA matrix
dist_cal_ADT	similarity metrics used for ADT matrix
ADT_subset	A vector indicates the subset that will be used.
K_knn	Number of nearest neighbours
K_knn_Aff	Number of nearest neighbors for computing affinity matrix
sigma	Variance for local model for computing affinity matrix
t	Number of iterations for the diffusion process.
metadata_names	A vector indicates the names of metadata returned
verbose	whether print out the process
topN	top highly variable genes are used variable gene selection (see ‘modelGeneVar’ in ‘scran’ package for more details)

**Value**

A SingleCellExperiment object with fused matrix results stored

**References**

B Wang, A Mezlini, F Demir, M Fiume, T Zu, M Brudno, B Haibe-Kains, A Goldenberg (2014) Similarity Network Fusion: a fast and effective method to aggregate multiple data types on a genome wide scale. Nature Methods. Online. Jan 26, 2014

**Examples**

```
data("sce_ctcl_subset", package = "CiteFuse")
sce_ctcl_subset <- CiteFuse(sce_ctcl_subset)
```

---

CITEseq_example	<i>A subset of ECCITE-seq data (control)</i>
-----------------	--

---

**Description**

Data from Mimitou et al. ECCITE-seq PBMC control sample data, which is a list of three matrices of RNA, ADT and HTO

**Usage**

```
data(CITEseq_example, package = 'CiteFuse')
```

**Format**

An object of class `list` of length 3.

**Source**

Gene Expression Omnibus with the accession code GSE126310.

**References**

Mimitou, E. P., Cheng, A., Montalbano, A., et al. (2019). Multiplexed detection of proteins, transcriptomes, clonotypes and CRISPR perturbations in single cells. *Nature Methods*, 16(5), 409–412.

---

crossSampleDoublets	<i>crossSampleDoublets</i>
---------------------	----------------------------

---

**Description**

A function that perform normalisation for alternative expression

**Usage**

```
crossSampleDoublets(sce, altExp_name = NULL, totalExp_threshold = 10)
```

**Arguments**

sce                    A SingleCellExperiment object

altExp\_name         Name of alternative expression that will be used to perform normalisation. If it is NULL, it will set to HTO.

totalExp\_threshold         the threshold indicates for the HTO less than this threshold will be filtered from the analysis

**Value**

A SingleCellExperiment Object

**Examples**

```
data(CITEseq_example, package = "CiteFuse")
sce_citeseq <- preprocessing(CITEseq_example)
sce_citeseq <- normaliseExprs(sce = sce_citeseq,
altExp_name = "HTO",
transform = "log")
sce_citeseq <- crossSampleDoublets(sce_citeseq)
```

---

DEbubblePlot

*DEbubblePlot*

---

**Description**

A function to generate circlepack plot to visualise the marker for each cluster

**Usage**

```
DEbubblePlot(de_list)
```

**Arguments**

de\_list                 A list of results from 'DE genes ()'

**Value**

A ggplot to visualise the DE results via bubble plot

**Examples**

```

library(S4Vectors)
data(sce_control_subset, package = "CiteFuse")
sce_control_subset <- DEgenes(sce_control_subset,
  altExp_name = "none",
  group = sce_control_subset$SNF_W_louvain,
  return_all = TRUE,
  exprs_pct = 0.5)

sce_control_subset <- selectDEgenes(sce_control_subset,
  altExp_name = "none")

sce_control_subset <- DEgenes(sce_control_subset,
  altExp_name = "ADT",
  group = sce_control_subset$SNF_W_louvain,
  return_all = TRUE,
  exprs_pct = 0.5)

sce_control_subset <- selectDEgenes(sce_control_subset,
  altExp_name = "ADT")

rna_DEgenes <- metadata(sce_control_subset)[["DE_res_RNA_filter"]]
adt_DEgenes <- metadata(sce_control_subset)[["DE_res_ADT_filter"]]

rna_DEgenes <- lapply(rna_DEgenes, function(x){
  x$name <- gsub("hg19_", "", x$name)
  x})
DEbubblePlot(list(RNA = rna_DEgenes, ADT = adt_DEgenes))

```

---

DEcomparisonPlot

*DEcomparisonPlot*


---

**Description**

A function to visualise the pairwise comparison of pvalue in different data modality.

**Usage**

```
DEcomparisonPlot(de_list, feature_list)
```

**Arguments**

<code>de_list</code>	A list including two lists results from 'DE genes ()'.
<code>feature_list</code>	A list including two lists features indicating the selected subset of features will be visualised

**Value**

A ggplot2 to visualise the comparison plot of DE.

**Examples**

```
library(S4Vectors)
data(sce_control_subset)
sce_control_subset <- DEgenes(sce_control_subset,
  group = sce_control_subset$SNF_W_louvain,
  return_all = TRUE,
  exprs_pct = 0.5)

sce_control_subset <- selectDEgenes(sce_control_subset)

sce_control_subset <- DEgenes(sce_control_subset,
  altExp_name = "ADT",
  group = sce_control_subset$SNF_W_louvain,
  return_all = TRUE,
  exprs_pct = 0.5)

sce_control_subset <- selectDEgenes(sce_control_subset,
  altExp_name = "ADT")

rna_list <- c("hg19_CD4",
  "hg19_CD8A",
  "hg19_HLA-DRB1",
  "hg19_ITGAX",
  "hg19_NCAM1",
  "hg19_CD27",
  "hg19_CD19")

adt_list <- c("CD4", "CD8", "MHCII (HLA-DR)", "CD11c", "CD56", "CD27", "CD19")

rna_DEgenes_all <- S4Vectors::metadata(sce_control_subset)[["DE_res_RNA"]]
adt_DEgenes_all <- S4Vectors::metadata(sce_control_subset)[["DE_res_ADT"]]

feature_list <- list(RNA = rna_list, ADT = adt_list)
de_list <- list(RNA = rna_DEgenes_all, ADT = adt_DEgenes_all)

DEcomparisonPlot(de_list = de_list,
  feature_list = feature_list)
```

---

DEgenes

*DEgenes*


---

**Description**

A function to perform DE analysis on CITE seq data

**Usage**

```
DEgenes(
  sce,
  altExp_name = "none",
  exprs_value = "logcounts",
  group = NULL,
  method = "wilcox",
  exprs_pct = 0.1,
  exprs_threshold = 0,
  return_all = FALSE,
  pval_adj = 0.05,
  mean_diff = 0,
  pct_diff = 0.1,
  topN = 10
)
```

**Arguments**

sce	A SingleCellExperiment object
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
exprs_value	A character indicates which expression value in assayNames is used.
group	A vector indicates the grouping of the data
method	A character indicates the method used in DE analysis
exprs_pct	A numeric indicates the threshold expression percentage of a gene to be considered in DE analysis
exprs_threshold	A numeric indicates the threshold of expression. By default is 0.
return_all	Whether return full list of DE genes
pval_adj	A numeric indicates the threshold of adjusted p-value.
mean_diff	A numeric indicates the threshold of difference of average expression.
pct_diff	A numeric indicates the threshold of difference of percentage expression.
topN	A numeric indicates the top number of genes will be included in the list.

**Value**

A SingleCellExperiment with DE results stored in meta data DE\_res

**Examples**

```
data(sce_control_subset)
sce_control_subset <- DEgenes(sce_control_subset,
  group = sce_control_subset$SNF_W_louvain,
  return_all = TRUE,
  exprs_pct = 0.5)
```



```
sce_control_subset <- selectDEgenes(sce_control_subset)
```

---

DEgenesCross

*DEgenesCross*


---

## Description

A function to perform DE analysis on a list of CITE seq data

## Usage

```
DEgenesCross(
  sce_list,
  altExp_name = "none",
  exprs_value = "logcounts",
  method = "wilcox",
  colData_name = NULL,
  group_to_test = NULL,
  exprs_pct = 0.1,
  exprs_threshold = 0,
  return_all = FALSE,
  pval_adj = 0.05,
  mean_diff = 0,
  pct_diff = 0.1,
  topN = 10
)
```

## Arguments

sce_list	A Slist of ingleCellExperiment object
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
exprs_value	A character indicates which expression value in assayNames is used.
method	A character indicates the method used in DE analysis
colData_name	A vector of character indicates the colData that stored the group information of each sce of the sce_list
group_to_test	A vector of character indicates which group in each sce is used to compared across the sce list.
exprs_pct	A numeric indicates the threshold expression percentage of a gene to be considered in DE analysis
exprs_threshold	A numeric indicates the threshold of expression. By default is 0.
return_all	Whether return full list of DE genes
pval_adj	A numeric indicates the threshold of adjusted p-value.

mean\_diff      A numeric indicates the threshold of difference of average expression.  
 pct\_diff        A numeric indicates the threshold of difference of percentage expression.  
 topN            A numeric indicates the top number of genes will be included in the list.

**Value**

A SingleCellExperiment with DE results stored in meta data DE\_res

**Examples**

```
data("sce_control_subset", package = "CiteFuse")
data("sce_ctcl_subset", package = "CiteFuse")

de_res <- DEgenesCross(sce_list = list(control = sce_control_subset,
ctcl = sce_ctcl_subset),
colData_name = c("SNF_W_louvain", "SNF_W_louvain"),
group_to_test = c("2", "6"))
```

---

geneADTnetwork	<i>geneADTnetwork</i>
----------------	-----------------------

---

**Description**

A function to visualise the features distribtuion

**Usage**

```
geneADTnetwork(
  sce,
  RNA_exprs_value = "logcounts",
  altExp_name = "ADT",
  altExp_exprs_value = "logcounts",
  RNA_feature_subset = NULL,
  ADT_feature_subset = NULL,
  cell_subset = NULL,
  cor_threshold = 0.5,
  cor_method = c("pearson", "kendall", "spearman"),
  RNA_exprs_pct = 0.1,
  ADT_exprs_pct = 0.1,
  RNA_exprs_threshold = 0,
  ADT_exprs_threshold = 0,
  network_layout = NULL,
  return_igraph = FALSE
)
```

**Arguments**

sce	A singlecellexperiment object
RNA_exprs_value	A character indicates which expression value for RNA in assayNames is used.
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
altExp_exprs_value	A character indicates which expression value in assayNames is used.
RNA_feature_subset	A vector of characters indicates the subset of features of RNA that are used for visualisation
ADT_feature_subset	A vector of characters indicates the subset of features of ADT that are used for visualisation
cell_subset	A vector of characters indicates the subset of cells that are used for visualisation
cor_threshold	Thresholds of correlation.
cor_method	a character string indicating which correlation coefficient (or covariance) is to be computed. One of "pearson" (default), "kendall", or "spearman": can be abbreviated.
RNA_exprs_pct	A numeric indicates the threshold expression percentage of a gene to be considered in correlation analysis
ADT_exprs_pct	A numeric indicates the threshold expression percentage of a gene to be considered in correlation analysis
RNA_exprs_threshold	A numeric indicates the threshold of RNA expression. By default is 0.
ADT_exprs_threshold	A numeric indicates the threshold of ADT expression. By default is 0.
network_layout	layout of the network
return_igraph	indicates whether return the igraph object

**Value**

A igraph object of gene-ADT network

**Examples**

```
library(SingleCellExperiment)
set.seed(2020)
data(sce_control_subset, package = "CiteFuse")
RNA_feature_subset <- sample(rownames(sce_control_subset), 50)
ADT_feature_subset <- rownames(altExp(sce_control_subset, "ADT"))

geneADTnetwork(sce_control_subset,
               RNA_feature_subset = RNA_feature_subset,
               ADT_feature_subset = ADT_feature_subset,
               cor_method = "pearson",
```

```
network_layout = igraph::layout_with_fr)
```

---

```
igraphClustering      igraphClustering
```

---

### Description

A function to perform igraph clustering

### Usage

```
igraphClustering(  
  sce,  
  metadata = "SNF_W",  
  method = c("louvain", "leiden", "walktrap", "spinglass", "optimal", "leading_eigen",  
            "label_prop", "fast_greedy", "edge_betweenness"),  
  ...  
)
```

### Arguments

sce	A singlecellexperiment object
metadata	indicates the meta data name of affinity matrix to virsualise
method	A character indicates the method for finding communities from igraph. Default is louvain clustering.
...	Other inputs for the igraph functions

### Value

A vector indicates the membership (clustering) results

### Examples

```
data(sce_control_subset, package = "CiteFuse")  
sce_control_subset <- CiteFuse(sce_control_subset)  
SNF_W_louvain <- igraphClustering(sce_control_subset,  
method = "louvain")
```

---

importanceADT	<i>importanceADT</i>
---------------	----------------------

---

### Description

A function to calculate the importance score of ADT

### Usage

```
importanceADT(
  sce,
  altExp_name = "ADT",
  exprs_value = "logcounts",
  method = c("randomForest", "PCA"),
  group = NULL,
  subsample = TRUE,
  times = 10,
  prop = 0.8,
  k_pca = 5,
  remove_first_PC = TRUE,
  ...
)
```

### Arguments

sce	A singlecellexperiment object
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
exprs_value	A character indicates which expression value in assayNames is used.
method	A character indicates the method of ADT importance calculation, either randomForest or PCA
group	A vector indicates the grouping of the data (for random forest)
subsample	Whether perform subsampling (for random forest)
times	A numeric indicates the times of subsampling is performed (for random forest)
prop	A numeric indicates the proportion of cells are subsampled from the whole data (for random forest)
k_pca	Number of principal component will be used to calculate the loading scores (for PCA)
remove_first_PC	A logical input indicates whether the first component will be removed from calculation (for PCA).
...	other arguments to 'randomForest()' or 'prcomp()' function

**Details**

For random forest, the importance scores are based on features importance. For PCA, it implements the method proposed in Levin et al (based on the loading of features).

**Value**

A SingleCellExperiment object

**References**

Levine, J.H., Simonds, E.F., Bendall, S.C., Davis, K.L., El-ad, D.A., Tadmor, M.D., Litvin, O., Fienberg, H.G., Jager, A., Zunder, E.R. and Finck, R., 2015. Data-driven phenotypic dissection of AML reveals progenitor-like cells that correlate with prognosis. *Cell*, 162(1), pp.184-197.

**Examples**

```
data("sce_control_subset", package = "CiteFuse")
sce_control_subset <- importanceADT(sce_control_subset,
group = sce_control_subset$SNF_W_louvain,
subsample = TRUE)
```

---

ligandReceptorTest      *ligandReceptorTest*

---

**Description**

A function to perform ligand receptor analysis

**Usage**

```
ligandReceptorTest(
  sce,
  ligandReceptor_list,
  cluster,
  RNA_exprs_value = "minMax",
  use_alt_exp = TRUE,
  altExp_name = "ADT",
  altExp_exprs_value = "zi_minMax",
  num_permute = 1000,
  p_sig = 0.05
)
```

**Arguments**

sce	A singlecellexperiment object
ligandReceptor_list	A data.frame indicates the ligand receptor list
cluster	A vector indicates the cluster results
RNA_exprs_value	A character indicates which expression value for RNA in assayNames is used.
use_alt_exp	A logical vector indicates whether receptors expression will use alternative expression matrix to quantify.
altExp_name	A character indicates which expression matrix is used. by default is ADT .
altExp_exprs_value	A character indicates which expression value in assayNames is used.
num_permute	Number of permutation.
p_sig	A numeric indicates threshold of the pvalue significance

**Value**

A SingleCellExperiment object with ligand receptor results

**Examples**

```
data(lr_pair_subset, package = "CiteFuse")
data(sce_control_subset, package = "CiteFuse")

sce_control_subset <- normaliseExprs(sce = sce_control_subset,
altExp_name = "ADT",
transform = "zi_minMax")

sce_control_subset <- normaliseExprs(sce = sce_control_subset,
altExp_name = "none",
exprs_value = "logcounts",
transform = "minMax")

sce_control_subset <- ligandReceptorTest(sce = sce_control_subset,
ligandReceptor_list = lr_pair_subset,
cluster = sce_control_subset$SNF_W_louvain,
RNA_exprs_value = "minMax",
use_alt_exp = TRUE,
altExp_name = "ADT",
altExp_exprs_value = "zi_minMax",
num_permute = 100)
```

---

lr_pair_subset	<i>A subset of Ligand Receptor Pairs</i>
----------------	--

---

**Description**

A subset of Ligand Receptor Pairs

**Usage**

```
data(lr_pair_subset, package = 'CiteFuse')
```

**Format**

An object of class matrix (inherits from array) with 50 rows and 2 columns.

---

normaliseExprs	<i>normaliseExprs</i>
----------------	-----------------------

---

**Description**

A function that perform normalisation for alternative expression

**Usage**

```
normaliseExprs(
  sce,
  altExp_name = NULL,
  exprs_value = "counts",
  transform = c("log", "clr", "zi_minMax", "minMax"),
  log_offset = NULL
)
```

**Arguments**

sce	A SingleCellExperiment object
altExp_name	Name of alternative expression that will be used to perform normalisation
exprs_value	A character indicates which expression value in assayNames is used.
transform	type of transformation, either log or clr (Centered log ratio transform)
log_offset	Numeric scalar specifying the pseudo-count to add when log-transforming expression values. Default is 1

**Value**

a SingleCellExperiment object



**Examples**

```
data(CITEseq_example, package = "CiteFuse")
sce_citeseq <- preprocessing(CITEseq_example)
sce_citeseq <- normaliseExprs(sce = sce_citeseq,
altExp_name = "ADT",
transform = "log")
```

---

plotHTO

*plotHTO*

---

**Description**

A function to plot HTO expression

**Usage**

```
plotHTO(sce, which_idx = seq_len(2), altExp_name = NULL, ncol = 2)
```

**Arguments**

sce	sce
which_idx	which_idx
altExp_name	altExp_name
ncol	ncol

**Value**

A plot visualising the HTO expression

**Examples**

```
data(CITEseq_example, package = "CiteFuse")
sce_citeseq <- preprocessing(CITEseq_example)
sce_citeseq <- normaliseExprs(sce = sce_citeseq,
altExp_name = "HTO",
transform = "log")
plotHTO(sce_citeseq, 1:4)
```

---

plotHTOSingle	<i>plotHTOSingle</i>
---------------	----------------------

---

**Description**

A function to plot HTO expression

**Usage**

```
plotHTOSingle(sce, which_idx = seq_len(2), altExp_name = NULL)
```

**Arguments**

sce	sce
which_idx	which_idx
altExp_name	altExp_name

**Value**

A plot visualising the HTO expression

---

preprocessing	<i>A function to preprocess the list of expression matrix</i>
---------------	---

---

**Description**

This function will keep the samples that are common across the list of expression matrix, and filter the features that are all zeros across samples, and finally construct a SingleCellExperiment object

**Usage**

```
preprocessing(
  exprsMat = NULL,
  return_sce = TRUE,
  assay_matrix = 1,
  filter_features = TRUE,
  rowData = NULL,
  colData = NULL
)
```

**Arguments**

<code>exprsMat</code>	A list or a matrix indicates the expression matrices of the testing datasets (each matrix must be <code>matrix</code> or <code>dgCMatrx</code> class)
<code>return_sce</code>	A logical input indicates whether a <code>SingleCellExperiment</code> object will be returned
<code>assay_matrix</code>	A integer indicates which list will be used as ‘assay’ input of ‘ <code>SingleCellExperiment</code> ’
<code>filter_features</code>	A logical input indicates whether the features with all zeros will be removed
<code>rowData</code>	A <code>DataFrame</code> indicates the <code>rowData</code> to be stored in the <code>sce</code> object
<code>colData</code>	A <code>DataFrame</code> indicates the <code>colData</code> to be stored in the <code>sce</code> object

**Value**

either a `SingleCellExperiment` object or a preprocessed expression matrix

**Examples**

```
data(CITEseq_example, package = "CiteFuse")
sce_citeseq <- preprocessing(CITEseq_example)
```

---

<code>readFrom10X</code>	<i>readFrom10X</i>
--------------------------	--------------------

---

**Description**

A function to read the data from 10X

**Usage**

```
readFrom10X(
  dir,
  type = c("auto", "sparse", "HDF5"),
  feature_named_by = c("gene_id", "gene_symbol"),
  filter_features = TRUE
)
```

**Arguments**

<code>dir</code>	A character indicates the directory of the 10X files
<code>type</code>	A character indicates the format of the data, <code>sparse</code> or <code>HDF5</code>
<code>feature_named_by</code>	A character indicates whether the genes will be named by <code>gene_id</code> or <code>gene_symbol</code>
<code>filter_features</code>	A logical input indicates whether the features with all zeros will be removed

**Value**

a SingleCellExperiment object

**Examples**

```
## Not run:
tmpdir <- tempdir()
tenXdata <- "http://cf.10xgenomics.com/samples/cell-exp/3.1.0/connect_5k_pbmc_NGSC3_ch1/"
file <- "connect_5k_pbmc_NGSC3_ch1_filtered_feature_bc_matrix.tar.gz"
download.file(paste0(tenXdata, file), file.path(tmpdir, file))
untar(file.path(tmpdir, file),
      exdir = tmpdir)
sce_citeseq_10X <- readFrom10X(file.path(tmpdir,
"filtered_feature_bc_matrix/"))
sce_citeseq_10X

## End(Not run)
```

---

reducedDimSNF

*reducedDimSNF*


---

**Description**

A function to reduce the dimension of the similarity matrix

**Usage**

```
reducedDimSNF(sce, metadata = "SNF_W", method = "UMAP", dimNames = NULL, ...)
```

**Arguments**

sce	A singlecellexperiment object
metadata	indicates the meta data name of affinity matrix to visualise
method	the method of visualisation, which can be UMAP, tSNE and diffusion map
dimNames	indicates the name of the reduced dimension results.
...	other parameters for tsne(), umap()

**Value**

A SingleCellExperiment object

**Examples**

```
data(sce_control_subset, package = "CiteFuse")
sce_control_subset <- CiteFuse(sce_control_subset)
sce_control_subset <- reducedDimSNF(sce_control_subset,
method = "tSNE",
dimNames = "tSNE_joint")
```

---

sce\_control\_subset      *A SingleCellExperiment of ECCITE-seq data*

---

**Description**

Data from Mimitou et al. ECCITE-seq PBMC Control sample data

**Usage**

```
data(sce_control_subset, package = 'CiteFuse')
```

**Format**

An object of class `SingleCellExperiment` with 1508 rows and 128 columns.

**Source**

Gene Expression Omnibus with the accession code GSE126310.

**References**

Mimitou, E. P., Cheng, A., Montalbano, A., et al. (2019). Multiplexed detection of proteins, transcriptomes, clonotypes and CRISPR perturbations in single cells. *Nature Methods*, 16(5), 409–412.

---

sce\_ctcl\_subset      *A SingleCellExperiment of ECCITE-seq data*

---

**Description**

Data from Mimitou et al. ECCITE-seq PBMC CTCL sample data

**Usage**

```
data(sce_ctcl_subset, package = 'CiteFuse')
```

**Format**

An object of class `SingleCellExperiment` with 1450 rows and 173 columns.

**Source**

Gene Expression Omnibus with the accession code GSE126310.

**References**

Mimitou, E. P., Cheng, A., Montalbano, A., et al. (2019). Multiplexed detection of proteins, transcriptomes, clonotypes and CRISPR perturbations in single cells. *Nature Methods*, 16(5), 409–412.

---

selectDEgenes	<i>selectDEgenes</i>
---------------	----------------------

---

**Description**

A function to select DE genes

**Usage**

```
selectDEgenes(
  sce = NULL,
  de_res = NULL,
  altExp_name = "none",
  pval_adj = 0.05,
  mean_diff = 0,
  pct_diff = 0.1,
  topN = 10
)
```

**Arguments**

sce	A SingleCellExperiment object with DE results stored in meta data DE_res list.
de_res	DE_res returned by DEgenesCross().
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
pval_adj	A numeric indicates the threshold of adjusted p-value.
mean_diff	A numeric indicates the threshold of difference of average expression.
pct_diff	A numeric indicates the threshold of difference of percentage expression.
topN	A numeric indicates the top number of genes will be included in the list.

**Value**

A SingleCellExperiment With filtered DE results in DE\_res\_filter list of metadata

## Examples

```
data(sce_control_subset)
sce_control_subset <- DEgenes(sce_control_subset,
group = sce_control_subset$SNF_W_louvain,
return_all = TRUE,
exprs_pct = 0.5)

sce_control_subset <- selectDEgenes(sce_control_subset)
```

---

spectralClustering      *spectralClustering*

---

## Description

A function to perform spectral clustering

## Usage

```
spectralClustering(affinity, K = 20, delta = 1e-05)
```

## Arguments

affinity	An affinity matrix
K	number of clusters
delta	delta

## Value

A list indicates the spectral clustering results

## Examples

```
data(sce_control_subset, package = "CiteFuse")
sce_control_subset <- CiteFuse(sce_control_subset)
SNF_W <- S4Vectors::metadata(sce_control_subset)[["SNF_W"]]
SNF_W_clust <- spectralClustering(SNF_W, K = 5)
```

---

visImportance	<i>visImportance</i>
---------------	----------------------

---

## Description

A function to visualise the features distribution

## Usage

```
visImportance(  
  sce,  
  plot = c("boxplot", "heatmap"),  
  altExp_name = "ADT",  
  exprs_value = "logcounts"  
)
```

## Arguments

sce	A singlecellexperiment object
plot	A string indicates the type of the plot (either boxplot or heatmap)
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
exprs_value	A character indicates which expression value in assayNames is used.

## Value

A plot (either ggplot or pheatmap) to visualise the ADT importance results

## Examples

```
data("sce_control_subset", package = "CiteFuse")  
sce_control_subset <- importanceADT(sce_control_subset,  
  group = sce_control_subset$SNF_W_louvain,  
  subsample = TRUE)  
visImportance(sce_control_subset, plot = "boxplot")
```



---

visLigandReceptor      *visLigandReceptor*

---

## Description

A function to visualise ligand receptor analysis

## Usage

```
visLigandReceptor(  
  sce,  
  type = c("pval_heatmap", "pval_dotplot", "group_network", "group_heatmap",  
           "lr_network"),  
  receptor_type = NULL  
)
```

## Arguments

sce	A singlecellexperiment object
type	A character indicates the type of the plot for ligand receptor results visualisation, option includes "pval_heatmap", "pval_dotplot", "group_network", "group_heatmap", and "lr_network"
receptor_type	A character indicates which receptor expression's ligand receptor results are used to generate the figures.

## Value

A plot visualise the ligand receptor results

## Examples

```
data(lr_pair_subset, package = "CiteFuse")  
data(sce_control_subset, package = "CiteFuse")  
  
sce_control_subset <- normaliseExprs(sce = sce_control_subset,  
  altExp_name = "ADT",  
  transform = "zi_minMax")  
  
sce_control_subset <- normaliseExprs(sce = sce_control_subset,  
  altExp_name = "none",  
  exprs_value = "logcounts",  
  transform = "minMax")  
  
sce_control_subset <- ligandReceptorTest(sce = sce_control_subset,  
  ligandReceptor_list = lr_pair_subset,  
  cluster = sce_control_subset$SNF_W_louvain,  
  RNA_exprs_value = "minMax",  
  use_alt_exp = TRUE,
```

```

                                altExp_name = "ADT",
                                altExp_exprs_value = "zi_minMax",
                                num_permute = 100)
visLigandReceptor(sce_control_subset,
type = "pval_heatmap",
receptor_type = "ADT")

```

---

visualiseDim

*visualiseDim*


---

## Description

A function to visualise the reduced dimension

## Usage

```

visualiseDim(
  sce,
  dimNames = NULL,
  colour_by = NULL,
  shape_by = NULL,
  data_from = c("colData", "assay", "altExp"),
  assay_name = NULL,
  altExp_name = NULL,
  altExp_assay_name = NULL,
  dim = seq_len(2)
)

```

## Arguments

sce	A singlecellexperiment object
dimNames	indicates the name of the reduced dimension results.
colour_by	A character indicates how the cells coloured by. The information either stored in colData, assay, or altExp.
shape_by	A character indicates how the cells shaped by. The information either stored in colData, assay, or altExp.
data_from	A character indicates where the colour by data stored
assay_name	A character indicates the assay name of the expression
altExp_name	A character indicates the name of alternative expression
altExp_assay_name	A character indicates the assay name of alternative expression
dim	a vector of numeric with length of 2 indicates which component is being plot

## Value

A ggplot of the reduced dimension visualisation

**Examples**

```

data(sce_control_subset, package = "CiteFuse")
sce_control_subset <- CiteFuse(sce_control_subset)
sce_control_subset <- reducedDimSNF(sce_control_subset,
method = "tSNE",
dimNames = "tSNE_joint")
visualiseDim(sce_control_subset, dimNames = "tSNE_joint",
colour_by = "SNF_W_clust")

```

---

visualiseExprs

*visualiseExprs*


---

**Description**

A function to visualise the features distribution

**Usage**

```

visualiseExprs(
  sce,
  plot = c("boxplot", "violin", "jitter", "density", "pairwise"),
  altExp_name = c("none"),
  exprs_value = "logcounts",
  group_by = NULL,
  facet_by = NULL,
  feature_subset = NULL,
  cell_subset = NULL,
  n = NULL,
  threshold = NULL
)

```

**Arguments**

sce	A singlecellexperiment object
plot	Type of plot, includes boxplot, violin, jitter, density, and pairwise. By default is boxplot
altExp_name	A character indicates which expression matrix is used. by default is none (i.e. RNA).
exprs_value	A character indicates which expression value in assayNames is used.
group_by	A character indicates how is the expression will be group in the plots (stored in colData).
facet_by	A character indicates how is the expression will be lay out panels in a grid in the plots (stored in colData).
feature_subset	A vector of characters indicates the subset of features that are used for visualisation

cell_subset	A vector of characters indicates the subset of cells that are used for visualisation
n	A numeric indicates the top expressed features to show.
threshold	Thresholds of high expression for features (only is used for pairwise plot).

**Value**

A ggplot to visualise te features distribution

**Examples**

```
data(sce_control_subset)
visualiseExprs(sce_control_subset,
plot = "boxplot",
group_by = "SNF_W_louvain",
feature_subset = c("hg19_CD8A"))

visualiseExprs(sce_control_subset,
plot = "density",
altExp_name = "ADT",
group_by = "SNF_W_louvain",
feature_subset = c("CD8", "CD4"))
```

---

visualiseExprsList      *visualiseExprsList*

---

**Description**

A function to visualise the features distribtuion for a list of SingleCellExperiment

**Usage**

```
visualiseExprsList(
  sce_list,
  plot = c("boxplot", "violin", "jitter", "density"),
  altExp_name = "none",
  exprs_value = "logcounts",
  group_by = NULL,
  feature_subset = NULL,
  cell_subset = NULL,
  n = NULL
)
```

**Arguments**

<code>sce_list</code>	A list of SingleCellExperiment object
<code>plot</code>	Type of plot, includes boxplot, violin, jitter, density, and pairwise. By default is boxplot
<code>altExp_name</code>	A character indicates which expression matrix is used. by default is none (i.e. RNA).
<code>exprs_value</code>	A character indicates which expression value in assayNames is used.
<code>group_by</code>	A character indicates how is the expression will be group in the plots (stored in colData).
<code>feature_subset</code>	A vector of characters indicates the subset of features that are used for visualisation
<code>cell_subset</code>	A vector of characters indicates the subset of cells that are used for visualisation
<code>n</code>	A numeric indicates the top expressed features to show.

**Value**

A ggplot to visualise te features distribution

**Examples**

```
data(sce_control_subset, package = "CiteFuse")
data(sce_ctcl_subset, package = "CiteFuse")
visualiseExprsList(sce_list = list(control = sce_control_subset,
ctcl = sce_ctcl_subset),
plot = "boxplot",
altExp_name = "none",
exprs_value = "logcounts",
feature_subset = c("hg19_CD8A"),
group_by = c("SNF_W_louvain", "SNF_W_louvain"))
```

---

visualiseKNN

*visualiseKNN*

---

**Description**

A function to perform louvain clustering

**Usage**

```
visualiseKNN(sce, colour_by = NULL, metadata = "SNF_W")
```

**Arguments**

sce	A singlecellexperiment object
colour_by	the name of coldata that is used to colour the node
metadata	indicates the meta data name of affinity matrix to virsualise

**Value**

A igraph plot

**Examples**

```
data(sce_control_subset, package = "CiteFuse")
sce_control_subset <- CiteFuse(sce_control_subset)
SNF_W_louvain <- igraphClustering(sce_control_subset,
method = "louvain")
visualiseKNN(sce_control_subset, colour_by = "SNF_W_louvain")
```

---

`withinSampleDoublets` *withinSampleDoublets*

---

**Description**

doublet identification within batch

**Usage**

```
withinSampleDoublets(sce, altExp_name = NULL, eps = 200, minPts = 50)
```

**Arguments**

sce	a SingleCellExperiment
altExp_name	expression name of HTO matrix
eps	eps of DBSCAN
minPts	minPts of DBSCAN

**Value**

A SingleCellExperiment object

**Examples**

```
data(CITEseq_example, package = "CiteFuse")
sce_citeseq <- preprocessing(CITEseq_example)
sce_citeseq <- normaliseExprs(sce = sce_citeseq,
altExp_name = "HT0",
transform = "log")
sce_citeseq <- crossSampleDoublets(sce_citeseq)
sce_citeseq <- withinSampleDoublets(sce_citeseq,
minPts = 10)
```

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