# Package 'bgx'

May 14, 2024

Title Bayesian Gene eXpression

**Version** 1.70.0

Author Ernest Turro, Graeme Ambler, Anne-Mette K Hein
Maintainer Ernest Turro <et341@cam.ac.uk></et341@cam.ac.uk>
<b>Description</b> Bayesian integrated analysis of Affymetrix GeneChips
License GPL-2
<b>Depends</b> R (>= 2.0.1), Biobase, affy (>= 1.5.0), gcrma (>= 2.4.1)
Suggests affydata, hgu95av2cdf
biocViews Microarray, DifferentialExpression
<b>Imports</b> Rcpp (>= 0.11.0)
LinkingTo Rcpp
git_url https://git.bioconductor.org/packages/bgx
git_branch RELEASE_3_19
git_last_commit 83539c4
git_last_commit_date 2024-04-30
Repository Bioconductor 3.19
Date/Publication 2024-05-14
Contents
analysis.bgx
bgx
mcmc.bgx
readOutput.bgx
saveAffinityPlot.bgx
setupVars.bgx
Index 10

2 analysis.bgx

|--|

## Description

Functions for plotting expression densities, differential expression densities, histogram of proportion of differentially expressed genes, etc.

#### **Usage**

```
plotExpressionDensity(bgxOutput, gene=NULL, normalize=c("none", "mean", "loess"),...)
plotDEDensity(bgxOutput, gene=NULL, conditions=c(1,2), normalize=c("none", "mean", "loess"), normgene
plotDEHistogram(bgxOutput, conditions=c(1,2), normalize=c("none", "mean", "loess"), normgenes=c(1:ler
rankByDE(bgxOutput, conditions=c(1,2), normalize=c("none", "mean", "loess"), normgenes=c(1:length(bg
plotDiffRank(bgxOutput, conditions=c(1,2), normalize=c("none", "mean", "loess"), normgenes=c(1:length)
```

## **Arguments**

bgx0utput	A list obtained from running readOutput.bgx on a BGX output directory.
gene	Which gene to analyse. This can either be an index or a name.
conditions	Indices of conditions to compare.
normalize	"none": do not normalise posterior distributions of mu. "mean": normalise by scaling posterior distributions of mu for conditions > 1 to have the same mean as the posterior distribution of mu for condition 1. "loess": same as "mean" but use loess normalisation.
normgenes	Which genes to use for loess normalisation. By default, use all genes.
df	Residual degrees of freedom. Decrease to 6 if the histogram fit goes haywire.
absolute	Rank genes by absolute differential expression.
ymax	Specify upper limit of y axis.
	Parameters to pass to density function (where applicable).

## Details

plotExpressionDensity plots gene expression distributions under various conditions for the specified gene.

plotDEDensity plots the differential expression distribution between two conditions for a given gene.

plotDEHistogram plots a histogram of differential expression between two conditions and estimates the number of up and down regulated differentially expressed genes.

rankByDE ranks genes by differential expression and returns ordering and corresponding DE values in a matrix.

plotDiffRank plots 2.5-97.5% confidence intervals for ranked differential expression estimates.

bgx 3

### Value

None, except plotDERank, which returns a matrix of genes ranked by differential expression.

#### Author(s)

Ernest Turro

### See Also

bgx, standalone.bgx, readOutput.bgx, plotExpressionDensity, plotDEDensity, plotDEHistogram

bgx	Fully Bayesian integrated approach to the analysis of Affymetrix GeneChip data

## **Description**

'bgx' estimates Bayesian Gene eXpression (BGX) measures from an AffyBatch object.

'standalone.bgx' creates various files needed by the bgx standalone binary and places them in a directory. One of these files is 'infile.txt'. In order to run standalone BGX, compile it and run 'bgx <path\\_to\\_infile.txt>' from the command line.

## Usage

```
bgx(aData, samplesets = NULL, genes = NULL, genesToWatch = NULL,
  burnin = 8192, iter = 16384, output = c("minimal","trace","all"),
  probeAff = TRUE, probecat_threshold = 100, adaptive = TRUE, rundir = ".")

standalone.bgx(aData, samplesets = NULL, genes = NULL, genesToWatch = NULL,
  burnin = 8192, iter = 16384, output = c("minimal", "trace", "all"),
  probeAff = TRUE, probecat_threshold = 100,
  adaptive = TRUE, batch_size = 50, optimalAR = 0.44, inputdir = "input")
```

## **Arguments**

aData	An AffyBatch object.
samplesets	A numeric vector specifying which condition each array belongs to. E.g. if samplesets=c(2,2), then the first two replicates belong to one condition and the last two replicates belong to another condition. If NULL, each array is assumed to belong to a different condition. If the aData object contains information about the experiment design in its phenoData slot, this argument is not required.
genes	A numeric vector specifying which genes to analyse. If NULL, all genes are analysed.
genesToWatch	A numeric vector specifying which genes to monitor closely amongst those chosen to be analysed (see below for details).

4 bgx

burnin Number of burn-in iterations.

iter Number of post burn-in iterations.

output One of "minimal", "trace" or "all". See below for details.

probeAff Stratify the mean (lambda) of the cross-hybridisation parameter (H) by cate-

gories according to probe-level sequence information.

probecat\_threshold

Minimum amount of probes per probe affinity category.

adaptive Adapt the variance of the proposals for Metropolis Hastings objects (that is: S,

H, Lambda, Eta, Sigma and Mu).

batch\_size Size of batches for calculating acceptance ratios and updating jumps.

optimalAR Optimal acceptance ratio.

rundir The directory in which to save the output runs.

inputdir The name of the directory in which to place the input files for the standalone

binary.

#### **Details**

• genesToWatchSpecify the subset of genes for which thinned samples from the full posterior distributions of log(S+1) (x) and log(H+1) (y) are collected.

- outputOutput the following to disk:
  - "minimal"The gene expression measure (muave), thinned samples from the full posterior distributions of mu (mu.[1..c]), where 'c' is the number of conditions, the integrated autocorrelation time (IACT) and the Markov chain Monte Carlo Standard Error (MCSE) for each gene under each condition. Note that the IACT and MCSE are calculated from the thinned samples of mu.
  - "trace"The same as "minimal" plus thinned samples from the full posterior distributions of sigma2 (sigma2.[1..c]), lambda (lambda.[1..s]), eta2 (eta2), phi (phi) and tau2 (tau2), where 's' is the number of samples. If there are probes with unknown sequences, output a thinned trace of their categorisation.
  - "all"The same as "trace" plus acceptance ratios for S (sacc), H (hacc), mu (muacc), sigma (sigmaacc), eta (etaacc) and lambda (lambdasacc).

## Value

'bgx' returns an ExpressionSet object containing gene expression information for each gene under each condition (not each replicate).

'standalone.bgx' returns the path to the BGX input files.

#### Note

The bgx() method and the bgx standalone binary create a directory in the working directory called 'run.x' (x:1,2,3,...), wherein files are placed for further detailed analysis.

## Author(s)

Ernest Turro

mcmc.bgx 5

#### References

Turro, E., Bochkina, N., Hein, A., Richardson, S. (2007) BGX: a Bioconductor package for the Bayesian integrated analysis of Affymetrix GeneChips. BMC Bioinformatics 2007, 8:439.

Hein, A., Richardson, S. (2006) A powerful method for detecting differentially expressed genes from GeneChip arrays that does not require replicates. BMC Bioinformatics 2006, 7:353.

Hein, A., Richardson, S., Causton, H., Ambler, G., Green., P. (2005) BGX: a fully Bayesian integrated approach to the analysis of Affymetrix GeneChip data. Biostatistics, 6, 3, pp. 349-373.

Hekstra, D., Taussig, A. R., Magnasco, M., and Naef, F. (2003) Absolute mRNA concentrations from sequence-specific calibration of oligonucleotide array. Nucleic Acids Research, 31. 1962-1968.

G.O. Roberts, J.S. Rosenthal (September, 2006) Examples of Adaptive MCMC.

#### **Examples**

```
# This example requires the 'affydata' and 'hgu95av2cdf' packages
if(require(affydata) && require(hgu95av2cdf)) {
  data(Dilution)
  eset <- bgx(Dilution, samplesets=c(2,2), probeAff=FALSE, burnin=4096, iter=8192,
      genes=c(12500:12599), output="all", rundir=file.path(tempdir()))
}</pre>
```

mcmc.bgx

Internal wrapper function for calling the bgx C++ function.

## **Description**

This internal function calls the bgx method in a loaded bgx shared object (bgx.so/bgx.dll)

## Usage

```
mcmc.bgx(pm, mm, samplesets, probesets, numberCategories, categories, unknownProbeSeqs, numberOfUnknown numberGenesToWatch, whichGenesToWatch, whichProbesToWatch, iter, burnin, adaptive, batch_size=50, optimalAR=0.44, output, samplenames = "unknown", subsample = ifelse(iter > 1024, iter/1024, 1), seed = 192492, rundir)
```

### **Arguments**

pm Perfect Match probes

mm MisMatch probes

samplesets A numeric vector specifying which condition each array belongs to. E.g. if samplesets=c(2,2), then the first two replicates belong to one condition and the last two replicates belong to another condition. If NULL, each array is assumed to belong to a different condition.

probesets A numeric vector specifying how probes are grouped into probesets.

6 mcmc.bgx

numberCategories

Number of probe affinity categories.

categories A numeric vector specifying which category each probe belongs to.

unknownProbeSeqs

A numeric vector specifying which probes lack sequence information.

numberOfUnknownProbeSeqs

Number of probes lacking sequence information.

numberGenesToWatch

How many genes to monitor closely.

whichGenesToWatch

A numeric vector specifying which genes to monitor closely.

whichProbesToWatch

The starting position for each probe in each gene to monitor closely.

iter Number of post burn-in iterations.

burnin Number of burn-in iterations.

adaptive Use adaptive MCMC for better mixing.

batch\_size Batch size for adaptive MCMC.

optimalAR Optimal acceptance ratio.

output One of "minimal", "trace", "diagnostic" or "mcse".

samplenames Vector of names for each array.

subsample Subsampling interval. seed Seed for PRNG.

rundir The directory in which to place the output run directories.

#### **Details**

See bgx for more details.

## Value

The name of the output directory.

## Note

You shouldn't call this function directly, but if you do, make sure the appropriate shared object is loaded.

## Author(s)

Ernest Turro

#### See Also

bgx, standalone.bgx

readOutput.bgx 7

readOutput.bgx

Read in the output from a BGX run.

## **Description**

readOutput.bgx reads in output from BGX which can then be fed into BGX analysis functions.

## Usage

```
readOutput.bgx(...)
```

## **Arguments**

. . .

Paths of BGX output directories. If you specify more than one path, then the runs will be combined such that each condition from each run is treated as a different different from all the others.

## **Details**

See bgx for more details.

#### Value

A list containing data from the BGX output.

## Author(s)

Ernest Turro

## See Also

 $\verb|bgx|, \verb|standalone.bgx|, \verb|plotDED| ensity|, \verb|plotDED| ensity|, \verb|plotDEH| is to gram|$ 

```
saveAffinityPlot.bgx Save a plot of affinity categorisation.
```

## Description

This internal function saves a plot showing how probes were categorised into affinity categories.

## Usage

```
saveAffinityPlot.bgx(originalAffinities, categories, dir, probecat_threshold)
```

8 setupVars.bgx

#### **Arguments**

originalAffinities

The affinities of the probes.

categories The categories of the probes.

dir Name of a directory in which to save the plot.

probecat\_threshold

The minimum number of probes per category that was used to categorise the

probes.

#### Author(s)

Ernest Turro

#### References

See bgx

#### See Also

bgx

setupVars.bgx

Initialise variables needed to run BGX simulation.

## Description

This internal function initialises several variables, which it returns in a list.

### Usage

setupVars.bgx(data, samplesets, genes, genesToWatch, probeAff, probecat\_threshold, rounding\_dec\_place

### **Arguments**

data An AffyBatch object.

samplesets A numeric vector specifying which condition each array belongs to. E.g. if

samplesets=c(2,2), then the first two replicates belong to one condition and the last two replicates belong to another condition. If NULL, each array is assumed

to belong to a different condition.

genes A numeric vector specifying which genes to analyse. If NULL, all genes are

analysed.

genesToWatch A numeric vector specifying which genes to monitor closely amongst those cho-

sen to be analysed (see below for details).

probeAff Stratify the mean (lambda) for the cross-hybridisation parameter (H) by cate-

gories according to probe-level sequence information.

setupVars.bgx 9

probecat\_threshold

Minimum amount of probes per probe affinity category.

rounding\_dec\_places

The initial probe categorisation is done by rounding affinities to the nearest rounding\_dec\_places decimal places. 1 is a good value.

#### Value

A list:

pm Perfect Match probes.
mm MisMatch probes.

samplesets A numeric vector specifying which condition each array belongs to. E.g. if

samplesets=c(2,2), then the first two replicates belong to one condition and the last two replicates belong to another condition. If NULL, each array is assumed

to belong to a different condition.

probesets A numeric vector specifying how probes are grouped into probesets.

numberOfCategories

Number of probe affinity categories.

categories A numeric vector specifying which category each probe belongs to.

unknownProbeSeqs

A numeric vector specifying which probes lack sequence information.

numberOfUnknownProbeSeqs

Number of probes lacking sequence information.

genesToWatch A numeric vector specifying which genes to monitor closely.

firstProbeInEachGeneToWatch

The starting position for each probe in each gene to monitor closely.

numArrays Number of arrays.

### Note

This function shouldn't be called directly.

### Author(s)

**Ernest Turro** 

#### References

See bgx

#### See Also

bgx

## **Index**

```
* IO
    analysis.bgx, 2
    readOutput.bgx, 7
* internal
    mcmc.bgx, 5
    saveAffinityPlot.bgx, 7
    setupVars.bgx, 8
* manip
    bgx, 3
{\tt analysis.bgx, 2}
bgx, 3, 3, 6-9
\texttt{mcmc.bgx}, \textcolor{red}{5}
plotDEDensity, 3, 7
plotDEDensity (analysis.bgx), 2
plotDEHistogram, 3, 7
plotDEHistogram(analysis.bgx), 2
plotDiffRank (analysis.bgx), 2
plotExpressionDensity, 3, 7
plotExpressionDensity(analysis.bgx), 2
rankByDE (analysis.bgx), 2
readOutput.bgx, 2, 3, 7
saveAffinityPlot.bgx, 7
setupVars.bgx, 8
standalone.bgx, 3, 6, 7
standalone.bgx (bgx), 3
```