

Package ‘cleaver’

May 14, 2024

Version 1.42.0

Title Cleavage of Polypeptide Sequences

Maintainer Sebastian Gibb <mail@sebastiangibb.de>

Depends R (>= 3.0.0), methods, Biostrings (>= 1.29.8)

Imports S4Vectors, IRanges

Suggests testthat (>= 0.8), knitr, BiocStyle (>= 0.0.14), rmarkdown, BRAIN, UniProt.ws (>= 2.36.5)

Description In-silico cleavage of polypeptide sequences. The cleavage rules are taken from:
http://web.expasy.org/peptide_cutter/peptidecutter_enzymes.html

License GPL (>= 3)

URL <https://github.com/sgibb/cleaver/>

BugReports <https://github.com/sgibb/cleaver/issues/>

LazyLoad yes

VignetteBuilder knitr

biocViews Proteomics

git_url <https://git.bioconductor.org/packages/cleaver>

git_branch RELEASE_3_19

git_last_commit 868c186

git_last_commit_date 2024-04-30

Repository Bioconductor 3.19

Date/Publication 2024-05-14

Author Sebastian Gibb [aut, cre] (<<https://orcid.org/0000-0001-7406-4443>>)

Contents

cleaver-package	2
cleave-methods	2

Index	10
--------------	-----------

cleaver-package *Cleavage of polypeptide sequences*

Description

This package cleaves polypeptide sequences. It provides three functions: [cleave](#), [cleavageRanges](#) and [cleavageSites](#).

Details

The cleavage rules are taken from: https://web.expasy.org/peptide_cutter/peptidecutter_enzymes.html

Package: cleaver
License: GPL (>= 3)
URL: <https://github.com/sgibb/cleaver/>

Author(s)

Sebastian Gibb <mail@sebastiangibb.de>

References

<https://github.com/sgibb/cleaver/>

Gasteiger E., Hoogland C., Gattiker A., Duvaud S., Wilkins M.R., Appel R.D., Bairoch A.; "Protein Identification and Analysis Tools on the ExPASy Server". (In) John M. Walker (ed): The Proteomics Protocols Handbook, Humana Press (2005).

https://web.expasy.org/peptide_cutter/peptidecutter_enzymes.html

See Also

[cleave](#), [cleavageRanges](#) and [cleavageSites](#).

cleave-methods *Cleavage of polypeptide sequences*

Description

This functions cleave polypeptide sequences. Use [cleavageSites](#) to find the cleavage sites, [cleavageRanges](#) to find the cleavage ranges and [cleave](#) to get the cleavage products.

Usage

```
## S4 method for signature 'character'
cleave(x, enzym = "trypsin", missedCleavages = 0,
       custom = NULL, unique = TRUE)

## S4 method for signature 'AAString'
cleave(x, enzym = "trypsin", missedCleavages = 0,
       custom = NULL, unique = TRUE)

## S4 method for signature 'AAStringSet'
cleave(x, enzym = "trypsin", missedCleavages = 0,
       custom = NULL, unique = TRUE)

## S4 method for signature 'character'
cleavageRanges(x, enzym = "trypsin", missedCleavages = 0,
              custom = NULL)

## S4 method for signature 'AAString'
cleavageRanges(x, enzym = "trypsin", missedCleavages = 0,
              custom = NULL)

## S4 method for signature 'AAStringSet'
cleavageRanges(x, enzym = "trypsin", missedCleavages = 0,
              custom = NULL)

## S4 method for signature 'character'
cleavageSites(x, enzym = "trypsin", custom = NULL)

## S4 method for signature 'AAString'
cleavageSites(x, enzym = "trypsin", custom = NULL)

## S4 method for signature 'AAStringSet'
cleavageSites(x, enzym = "trypsin", custom = NULL)
```

Arguments

x	polypeptide sequences.
enzym	character, cleavage rule.
missedCleavages	numeric, number of missed cleavages.
custom	character, of length 1 or 2. Could be used to define own cleavage rules. The first element would be the pattern and the optional second element would be an exception (non-cleavage) pattern. Perl-like regular expressions are supported, see gregexpr for details. If custom is set the enzym is ignored.
unique	logical, if TRUE all duplicated cleavage products per peptide are removed.

Details

The cleavage rules are taken from: https://web.expasy.org/peptide_cutter/peptidecutter_enzymes.html

Cleavage rules (cleavage between P1 and P1'):

Rule name	P4	P3	P2	P1	P1'
arg-c proteinase	-	-	-	R	-
asp-n endopeptidase	-	-	-	-	D
bnps-skatole-c	-	-	-	W	-
caspase1	F,W,Y,L	-	H,A,T	D	not P,E,D,Q,K,R
caspase2	D	V	A	D	not P,E,D,Q,K,R
caspase3	D	M	Q	D	not P,E,D,Q,K,R
caspase4	L	E	V	D	not P,E,D,Q,K,R
caspase5	L,W	E	H	D	-
caspase6	V	E	H,I	D	not P,E,D,Q,K,R
caspase7	D	E	V	D	not P,E,D,Q,K,R
caspase8	I,L	E	T	D	not P,E,D,Q,K,R
caspase9	L	E	H	D	-
caspase10	I	E	A	D	-
chymotrypsin-high	-	-	-	F,Y	not P
	-	-	-	W	not M,P
chymotrypsin-low	-	-	-	F,L,Y	not P
	-	-	-	W	not M,P
	-	-	-	M	not P,Y
	-	-	-	H	not D,M,P,W
clostripain	-	-	-	R	-
cnbr	-	-	-	M	-
enterokinase	D,E	D,E	D,E	K	-
factor xa	A,F,G,I,L,T,V,M	D,E	G	R	-
formic acid	-	-	-	D	-
glutamyl endopeptidase	-	-	-	E	-
granzyme-b	I	E	P	D	-
hydroxylamine	-	-	-	N	G
iodosobenzoic acid	-	-	-	W	-
lysc	-	-	-	K	-
lysn	-	-	-	-	K
lysarginase	-	-	-	-	K,R
neutrophil elastase	-	-	-	A,V	-
ntcb	-	-	-	-	C
pepsin1.3	-	not H,K,R	not P	not R	F,L
pepsin	-	not H,K,R	not P	not R	F,L,W,Y
	-	not H,K,R	not P	F,L,W,Y	-
	-	not H,K,R	not P	F,L	-
proline endopeptidase	-	-	not H,K,R	P	not P
proteinase k	-	-	-	A,E,F,I,L,T,V,W,Y	-
staphylococcal peptidase i	-	-	not E	E	-
thermolysin	-	-	-	not D,E	A,F,I,L,M,V
thrombin	-	-	G	R	G

	A,F,G,I,L,T,V,M	A,F,G,I,L,T,V,W	P	R	not D,E
trypsin	-	-	-	K,R	not P
	-	-	W	K	P
	-	-	M	R	P
trypsin-high	-	-	-	K,R	not P
	-	-	W	K	P
	-	-	M	R	P
trypsin-low	-	-	-	K,R	not P
	-	-	W	K	P
	-	-	M	R	P
trypsin-simple	-	-	-	K,R	-

Exceptions:

Rule name	Enzyme name	P4	P3	P2	P1	P1'	P2'
trypsin	-	-	C,D	K	D	-	-
		-	-	C	K	H,Y	-
		-	-	C	R	K	-
		-	-	R	R	H,R	-
trypsin-high	-	-	C,D	K	D	-	-
		-	-	C	K	H,Y	-
		-	-	C	R	K	-
		-	-	R	R	H,R	-

Rule name	Enzyme name
arg-c proteinase	Arg-C proteinase
asp-n endopeptidase	Asp-N endopeptidase
bnps-skatole-c	BNPS-Skatole
caspace1	Caspase 1
caspace2	Caspase 2
caspace3	Caspase 3
caspace4	Caspase 4
caspace5	Caspase 5
caspace6	Caspase 6
caspace7	Caspase 7
caspace8	Caspase 8
caspace9	Caspase 9
caspace10	Caspase 10
chymotrypsin-high	Chymotrypsin-high specificity (C-term to [FYW], not before P)
chymotrypsin-low	Chymotrypsin-low specificity (C-term to [FYWML], not before P)
clostripain	Clostripain (Clostridiopeptidase B)
cnbr	CNBr
enterokinase	Enterokinase
factor xa	Factor Xa
formic acid	Formic acid

glutamyl endopeptidase	Glutamyl endopeptidase
granzyme-b	Granzyme B
hydroxylamine	Hydroxylamine
iodosobenzoic acid	Iodosobenzoic acid
lysc	LysC
lysn	LysN
lysarginase	LysargiNase
neutrophil elastase	Neutrophil elastase
ntcb	NTCB (2-nitro-5-thiocyanobenzoic acid)
pepsin1.3	Pepsin (pH == 1.3)
pepsin	Pepsin (pH > 2)
proline endopeptidase	Proline-endopeptidase
proteinase k	Proteinase K
staphylococcal peptidase i	Staphylococcal Peptidase I
thermolysin	Thermolysin
thrombin	Thrombin
trypsin	Trypsin
trypsin-high	Trypsin, higher specificity as defined in PeptideMass, identical to <i>trypsin</i>
trypsin-low	Trypsin, C-term to K/R if C-term is not P, as defined in PeptideMass
trypsin-simple	Trypsin, C-term to K/R, even before P, as defined in PeptideMass

Value

cleave If *x* is a character it returns a list of the same length as *x*. Each element contains a character vector with the corresponding cleavage products of the polypeptides. If *x* is an [AAString](#) or an [AAStringSet](#) an [AAStringSet](#) or an [AAStringSetList](#) instance of the same length as *x* is returned. Each element contains an [AAString](#) or an [AAStringSet](#) instance with the corresponding cleavage products of the polypeptides.

cleavageRanges If *x* is a character it returns a list of the same length as *x*. Each element contains a two-column matrix with the start and end positions of the peptides. If *x* is an [AAString](#) or an [AAStringSet](#) instance an [IRanges](#) or an [IRangesList](#) of the same length as *x* is returned.

cleavageSites Returns a list of the same length as *x*. Each element contains an integer vector with the cleavage positions.

Overview:

Input	cleave	cleavageRanges	cleavageSites
character	list of character	list of matrix	list of integer
AAString	AAStringSet	IRanges	list of integer
AAStringSet	AAStringSetList	IRangesList	list of integer

Author(s)

Sebastian Gibb <mail@sebastiangibb.de>

References

Gasteiger E., Hoogland C., Gattiker A., Duvaud S., Wilkins M.R., Appel R.D., Bairoch A.; "Protein Identification and Analysis Tools on the ExPASy Server". (In) John M. Walker (ed): The Proteomics Protocols Handbook, Humana Press (2005).

https://web.expasy.org/peptide_cutter/peptidecutter_enzymes.html

PeptideMass https://web.expasy.org/peptide_mass/peptide-mass-doc.html#table1

See Also

[AAString](#), [AAStringSet](#), [AAStringSetList](#), [IRanges](#), [IRangesList](#)

Examples

```
library("cleaver")

## Gastric juice peptide 1 (UniProtKB/Swiss-Prot: GAJU_HUMAN/P01358)
gaju <- "LAAGKVEDSD"

cleave(gaju, "trypsin")
# $LAAGKVEDSD
# [1] "LAAGK" "VEDSD"

cleavageRanges(gaju, "trypsin")
# $LAAGKVEDSD
#   start end
# [1,]    1  5
# [2,]    6 10

cleavageSites(gaju, "trypsin")
# $LAAGKVEDSD
# [1] 5

cleave(gaju, "trypsin", missedCleavages=1)
# $LAAGKVEDSD
# [1] "LAAGKVEDSD"

cleavageRanges(gaju, "trypsin", missedCleavages=1)
# $LAAGKVEDSD
#   start end
# [1,]    1 10

cleave(gaju, "trypsin", missedCleavages=0:1)
# $LAAGKVEDSD
# [1] "LAAGK" "VEDSD" "LAAGKVEDSD"

cleavageRanges(gaju, "trypsin", missedCleavages=0:1)
# $LAAGKVEDSD
#   start end
# [1,]    1  5
# [2,]    6 10
# [3,]    1 10
```

```

cleave(gaju, "pepsin")
# $LAAGKVEDSD
# [1] "LAAGKVEDSD"
# (no cleavage)

## use AAStrngSet
gaju <- AAStrngSet("LAAGKVEDSD")

cleave(gaju)
# AAStrngSetList of length 1
# [1] "LAAGKVEDSD" LAAGK VEDSD

## Beta-enolase (UniProtKB/Swiss-Prot: ENOB_THUAL/P86978)
enob <- "SITKIKAREILD"

cleave(enob, "trypsin")
# $SITKIKAREILD
# [1] "SITK" "IK" "AR" "EILD"

cleave(enob, "trypsin", missedCleavages=2)
# $SITKIKAREILD
# [1] "SITKIKAR" "IKAREILD"

cleave(enob, "trypsin", missedCleavages=0:2)
# $SITKIKAREILD
# [1] "SITK" "IK" "AR" "EILD" "SITKIK" "IKAR"
# [7] "AREILD" "SITKIKAR" "IKAREILD"

## define own cleavage rule: cleave at K
cleave(enob, custom="K")
# $SITKIKAREILD
# [1] "SITK" "IK" "AREILD"

cleavageRanges(enob, custom="K")
# $SITKIKAREILD
# start end
# [1,] 1 4
# [2,] 5 6
# [3,] 7 12

## define own cleavage rule: cleave at K but not if followed by A
cleave(enob, custom=c("K", "K(?=A)"))
# $SITKIKAREILD
# [1] "SITK" "IKAREILD"

cleavageRanges(enob, custom=c("K", "K(?=A)"))
# $SITKIKAREILD
# start end
# [1,] 1 4

```



```
# [2,]      5 12  
  
cleavageSites(enob, custom=c("K", "K(?=A)"))  
# $SITKIKAREILD  
# [1] 4
```

Index

* **methods**

cleave-methods, [2](#)

* **package**

cleaver-package, [2](#)

AAString, [6](#), [7](#)

AAStringSet, [6](#), [7](#)

AAStringSetList, [6](#), [7](#)

cleavageRanges, [2](#)

cleavageRanges (cleave-methods), [2](#)

cleavageRanges, AAString-method
(cleave-methods), [2](#)

cleavageRanges, AAStringSet-method
(cleave-methods), [2](#)

cleavageRanges, character-method
(cleave-methods), [2](#)

cleavageSites, [2](#)

cleavageSites (cleave-methods), [2](#)

cleavageSites, AAString-method
(cleave-methods), [2](#)

cleavageSites, AAStringSet-method
(cleave-methods), [2](#)

cleavageSites, character-method
(cleave-methods), [2](#)

cleave, [2](#)

cleave (cleave-methods), [2](#)

cleave, AAString-method
(cleave-methods), [2](#)

cleave, AAStringSet-method
(cleave-methods), [2](#)

cleave, character-method
(cleave-methods), [2](#)

cleave-methods, [2](#)

cleaver-package, [2](#)

gregexpr, [3](#)

IRanges, [6](#), [7](#)

IRangesList, [6](#), [7](#)